Formation of Superoxide Anion during Ferrous Ion–Induced Decomposition of Linoleic Acid Hydroperoxide under Aerobic Conditions

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Received May 7, 2003; accepted October 22, 2003

We studied the mechanism of formation of oxygen radicals during ferrous ion-induced decomposition of linoleic acid hydroperoxide using the spin trapping and chemiluminescence methods. The formation of the superoxide anion (O_2^{-}) was verified in the present study. The hydroxyl radical is also generated through Fenton type decomposition of hydrogen peroxide produced on disproportionation of O_2^{-} . A carbon-centered radical was detected using 5-(diethoxyphosphoryl)-5-methyl-1-pyrroline N-oxide (DEPMPO) as a spin trap. Alkoxyl radical formation is essential for the conversion of linoleic acid hydroperoxide into the peroxyl radical by ferrous ion. It is likely that the alkoxyl radical $[R_1CH(O)R_2]$ is converted into the hydroxylcarbon radical $[R_1C^{-}(OH)R_2]$ in water, and that this carbon radical reacts with oxygen to give the α -hydroxyperoxyl radical $[R_1R_2C(OH)OO]$, which decomposes into the carbocation $[R_1C^{+}(OH)R_2]$ and O_2^{-} .

Key words: ferrous ion-induced decomposition of linoleic acid hydroperoxide, hydrogen peroxide, hydroxyl radical, linoleic acid alkoxyl radical, superoxide anion.

Abbreviations: DEPMPO, 5-(diethoxyphosphoryl)-5-methyl-1-pyrroline N-oxide; DEPMPO-OH, DEPMPO-hydroxyl adduct; DEPMPO-OOH, superoxide anion adduct of DEPMPO; DMPO, 5,5-dimethyl-1-pyrroline N-oxide; DMPO-OH, DMPO-hydroxyl adduct; DMPO-OOH, superoxide anion adduct of DMPO; ESR, electron spin resonance; HPLC, high performance liquid chromatography; HX, hypoxanthine; LA, linoleic acid; LO, linoleic acid alkoxyl radical; LOH, linoleic acid hydroxide; LOO, linoleic acid peroxyl radical; LOOH, linoleic acid hydroxide; MCLA, 2-methyl-6-(p-methoxyphenyl)-3,7-dihydroimidazo[1,2-a]pyrazin-3-one; SOD, superoxide dismutase; XOD, xanthine oxidase.

The ferrous ion (Fe^{2+}) —catalyzed decomposition of unsaturated fatty acid hydroperoxides is generally accepted to occur as shown in equation 1, in which the alkoxyl type radical and hydoxyl anion are produced (1, 2).

$$LOOH + Fe^{2+} \rightarrow LO^{-} + OH^{-} + Fe^{3+}$$
 (1)

Using a spin trapping technique involving 5,5-dimethyl-1-pyrroline N-oxide (DMPO), however, Yagi $et\ al.$ found that Fe²⁺-induced decomposition of linoleic acid hydroper-oxide (LOOH) generates the hydroxyl radical (OH) under aerobic conditions (3). They suggested that Fe²⁺ causes homolytic cleavage of the O-O bond of LOOH (Eq. 2).

$$LOOH + Fe^{2+} \rightarrow LO^{\cdot} + OH + Fe^{2+}$$
 (2)

However, it is unlikely that Fe²⁺ induces the homolysis of LOOH. There is a possibility that a consecutive chain reaction helps to yield OH. Clarification of the reaction

*To whom correspondence should be addressed. Tel: +81-76-265-2217, Fax: +81-76-234-4233, E-mail: kambayashi@pub.m.kanazawa-u.ac.jp mechanism is important because the Fe²⁺-catalyzed decomposition of unsaturated fatty acid hydroperoxides is one of the most fundamental reactions in free radical biochemistry.

Schaich and Borg (4) and Chamulitrat et al. (5) also reported the formation of a spin adduct, which was comparable with the DMPO-OH adduct formed during the Fe²⁺-catalyzed decomposition of methyl linoleate hydroperoxide under aerobic conditions. However, they proposed a different mechanism from equation 1. The former claimed that the electron spin resonance (ESR) spectrum of the DMPO-lipid alkoxyl radical spin adduct was identical to that of DMPO-OH. The latter claimed that most DMPO-OH was formed through ferric ion (Fe³⁺)-catalyzed nucleophilic addition of water to DMPO (6), and/or by the reaction between DMPO and OH produced via the Fenton reaction (7). These interpretations are in conflict. In these studies, only DMPO was used as the trapping reagent. Obviously, an ESR study with only one spin trap will not establish any mechanisms. For example, DMPO-OH is formed in various ways, as mentioned below. The superoxide anion $(O_2$ -) adduct of DMPO (DMPO-OOH) is a relatively short-lived spin adduct and is reduced to

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DMPO-OH within a few min (8). DMPO-OH is also formed through Fe²⁺-catalayzed decomposition of DMPO-OOH (9). The reaction of singlet oxygen (10₂) with DMPO also gives DMPO-OH through a Fenton-like reaction (10, 11). Furthermore, DMPO-OH is produced under unexpected conditions such as on the Fe^{2+} - (12), Fe^{3+} - (6), or Fe3+-bleomycin complex- (13) catalyzed oxidation of DMPO, or the photooxidation of DMPO (12). This makes it difficult to determine how DMPO-OH is produced. Recently, 5-(diethoxyphosphoryl)-5-methyl-1-pyrroline N-oxide (DEPMPO) was developed as a new spin trap (14). The O₂ - spin adduct of DEPMPO (DEPMPO-OOH) is long-lived in comparison with DMPO-OOH (14, 15). Although DEPMPO-OH is observed in the presence of DEPMPO and Fe³⁺, a carbon-centered radical is also detected (16), unlike in the case where DMPO-OH is produced through Fe³⁺-catalyzed oxidation of DMPO. Therefore, DEPMPO is a useful spin trap for discriminating OH from O_2 in a solution in which the two species coexist.

The present study was undertaken to verify the oxygen radical formation mechanism during the Fe $^{2+}$ -catalyzed decomposition of LOOH under aerobic conditions, using the ESR spin trapping technique and the 2-methyl-6-(p-methoxyphenyl)-3,7-dihydroimidazo[1,2-a]pyrazin-3-one (MCLA)-dependent chemiluminescence methods. We used DEPMPO in addition to DMPO as a spin trap in this study.

MATERIALS AND METHODS

Materials—DMPO, lipoxidase (Type 1-B), superoxide dismutase (SOD, from bovine erythrocytes), catalase (from bovine liver), and uric acid were purchased from Sigma Chemical (St. Louis, MO). DEPMPO was obtained from OXIS International (Portland, OR). Ferrous sulfate (FeSO₄) heptahydrate and ferric chloride (FeCl₃) hexahydrate were purchased from Wako Pure Chemical Industries (Osaka). Linoleic acid (LA) and MCLA were obtained from Tokyo Chemical Industry (Tokyo). Solvents and other reagents were of the highest grade commercially available. LA was purified by high performance liquid chromatography (HPLC) before use. 13-Hydroperoxy-9-cis,11-trans-octadecadienoic acid (linoleic hydroperoxide; LOOH) was prepared using LA and lipoxidase, and LOOH was reduced to alcohol (LOH) with triphenylphosphine, and then the LOOH and LOH were purified by HPLC as previously described (17). The concentration of LOOH was determined using a hydroperoxide-specific, isoluminol chemiluminescence assay with HPLC (HPLC-CL) (18). The concentrations of LOOH and LOH were also determined using an assumed molar extinction coefficient at 234 nm, 28,000 M⁻¹ cm⁻¹ (19). DMPO was purified by filtration with decolorizing activated carbon (Aldrich Chemical Company, Milwaukee, WI) immediately before use (8, 20). The concentration of DMPO was determined using an assumed molar extinction coefficient at 234 nm, 7,700 M⁻¹ cm⁻¹ (21). DEPMPO was used without further purification. The concentration of DEPMPO was calculated from its molecular weight.

Spin Trapping of Free Radicals Produced during Fe²⁺-Catalyzed Decomposition of LOOH—A reaction mixture (450 µl) containing 2.5 mM LOOH and 613 mM DMPO

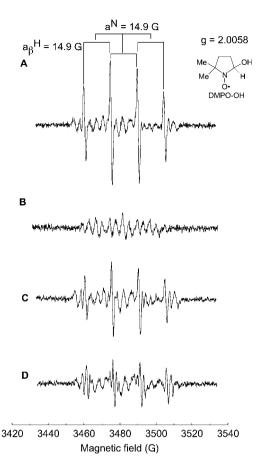
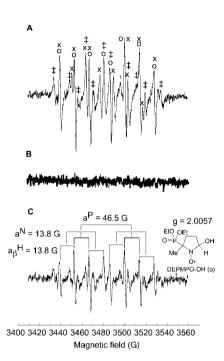
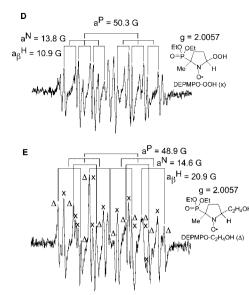


Fig. 1. ESR spectra of the DMPO-free radical spin adducts formed during the Fe²+-catalyzed decomposition of LOOH, and the effects of SOD and catalase thereon. A: 2.2 mM LOOH + 100 μ M Fe²+ + 613 mM DMPO, B: A – 2.2 mM LOOH, C: A + 0.5 μ M SOD, D: A + 50 μ g/ml catalase. Receiver gain: 2.5 × 10⁵; modulation frequency: 100 kHz; modulation amplitude: 1.011 G; conversion: 81.920 ms; time constant: 40.960 ms; sweep time: 83.883 s; microwave frequency: 9.779 GHz; microwave power: 7.73 mW; scan number: ×10.

was prepared in 50 mM potassium phosphate buffer (pH 7.4) pretreated with CHELEX 100 (BIO-RAD Laboratories, Hercules, CA). LOOH-decomposition was initiated by the addition of FeSO₄ (final concentration: 100 μM) at 25°C under aerobic conditions. LA or LOH was also used instead of LOOH. If necessary, SOD (0.5 µM), catalase (50 μg/ml), ethanol (16.7%), sodium azide (1 mM), or uric acid (100 µM) was added to the reaction mixture. FeCl₃ and DEPMPO (20 or 40 mM) were used instead of FeSO₄ and DMPO, respectively. An aliquot of the reaction mixture was transferred to a flat cell and ESR signals were measured with a BRUKER ESR spectrometer (ESP 300E. Karlsruhe, Germany) at 25°C. The measurement conditions are given in each figure legend. To avoid photo-induced reactions, sample preparation and measurements were carried out under weak red light. If necessary, argon gas was introduced into a solution for 2 h under a nitrogen atmosphere in a glove box prior to the measurement. A reaction solution was transferred to a flat cell. ESR signals were measured at 25°C under atmospheric air after the flat cell had been sealed with a teflon seal under a nitrogen atmosphere.





3400 3420 3440 3460 3480 3500 3520 3540 3560

Magnetic field (G)

Fig. 2. ESR spectra of the DEPMPOfree radical spin adducts formed during the Fe2+-catalyzed decomposition of LOOH, and the effects of SOD, catalase, and ethanol **thereon.** o, x, \ddagger , and Δ represent the DEPMPO-OH spin adduct, DEPMPO-OOH spin adduct, DEPMPO-carboncentered radical adduct, and DEPMPOhydroxyethyl spin adduct, respectively. A: $2.2 \text{ mM LOOH} + 100 \mu\text{M Fe}^{2+}$ + 40 mM DEPMPO, B: A - 2.2 mM LOOH, C: A + $0.5 \mu M$ SOD, D: A + 50μg/ml catalase, E: A + 16.7% ethanol. Receiver gain: 2.5 × 105; modulation frequency: 100 kHz; modulation amplitude: 1.011 G; conversion: 81.920 ms; time constant: 40.960 ms; sweep time: 83.883 s; microwave frequency: 9.779 GHz; microwave power: 23.9 mW; scan number: $\times 10$.

Chemiluminescence during Fe2+-Catalyzed Decomposition of LOOH—A reaction mixture (1 ml) comprising 50 μM LOOH and 10 μM MCLA in 50 mM potassium phosphate buffer (pH 7.4) pretreated with CHELEX 100 was placed in a luminescence reader (Type; BLR-301, Aloka, Tokyo). Chemiluminescence was detected after the addition of FeSO₄ (final concentration: 2 µM) at 25°C under aerobic conditions. If necessary, SOD (0.5 μM), catalase (20 μg/ml), sodium azide (1 mM), and/or DMPO (12 mM) were added to the reaction mixture. LA or LOH was also used instead of LOOH. Various concentrations of LA, LOH, LOOH, or FeSO₄ were also used, as described in the figure legends. If necessary, argon gas was introduced into a solution under atmospheric air for 30 min prior to the measurement. The effect of the FeSO₄/FeCl₃ (Fe = 2 μM) ratio on chemiluminescence intensity was also examined.

Yield of O_2 — during Fe^{2+} -Catalyzed Decomposition of LOOH—Hypoxanthine (HX; 1×10^{-3} U/ml, 5×10^{-3} U/ml, or 1×10^{-3} U/ml) and xanthine oxidase (XOD; $40~\mu M$) were incubated with cytochrome c (50 μM) at 25°C, the change in absorbance of cytochrome c at 550 nm being monitored. The amount of O_2 — formed was calculated using the molar extinction coefficient of a reduced form of cytochrome c at 550 nm [21,100 M⁻¹ cm⁻¹ (22)]. The values were compared with the integrated MCLA-dependent chemiluminescence intensity obtained under the same conditions except that MCLA (10 μM) was used instead of cytochrome c and the amount of O_2 — formed per chemiluminescence intensity was calculated.

Chemiluminescence intensity was measured during 2 μM Fe²+-catalyzed decomposition of 36 μM LOOH with 10 μM MCLA in the presence or absence of 0.5 μM SOD at 25°C. The change in LOOH was monitored by HPLC-CL during 2 μM Fe²+-induced decomposition of 36 μM LOOH, and the amount of LOOH decomposed during the present reaction was obtained. The yield of O_2 -- during Fe²+-catalyzed decomposition of LOOH was calculated with the

values obtained in the above experiments. $[O_2$ formed] was calculated from the SOD-sensitive MCLA-dependent chemiluminescence intensity.

Yield of
$$O_2^{\cdot-}$$
 (%) = $[O_2^{\cdot-} \text{ formed}] \ (\mu M)/[LOOH \ decreased] \ (\mu M) \times 100$

RESULTS

Spin Trapping of Free Radicals Produced during Fe²⁺-Catalyzed Decomposition of LOOH under Aerobic Conditions—Fig. 1A shows the ESR spectrum obtained during the Fe²⁺-induced decomposition of LOOH in the presence of DMPO. The hyperfine splitting constants of major signals were $a^{\rm N}=a_{\rm \beta}^{\rm \ \bar{H}}=14.9$ G, g=2.0058, which were identical with those of DMPO-OH ($a^{\text{N}} = a_{\beta}^{\text{H}} = 15.0 \text{ G}, g =$ 2.0062) (23). The DMPO-OH signal was not observed when LOOH was removed from the above system (Fig. 1B). The DMPO-OH signal intensity was decreased with the addition of SOD (Fig. 1C) and was substantially decreased on the addition of catalase (Fig. 1D). Chamulitrat et al. reported comparable results with catalase, although they used methyl linoleate hydroperoxide instead of LOOH (5). Separate experiments involving HPLC-CL showed that catalase (50 µg) did not decompose LOOH to the linoleic acid alkoxyl radical (LO) or linoleic acid peroxyl radical (LOO) (data not shown).

To obtain clearer ESR signals, we used DEPMPO as a spin trap instead of DMPO. Fig. 2A shows the ESR spectrum obtained during the Fe²+-catalyzed decomposition of LOOH in the presence of DEPMPO under aerobic conditions. The ESR spectrum consists of the signals of three kinds of spin adducts: the DEPMPO-hydroxyl adduct (DEPMPO-OH) (o; $a^{\rm N}$ = 14.0 G, $a_{\rm \beta}^{\rm H}$ = 13.0 G, $a^{\rm P}$ = 47.4 G, g = 2.0059) (14), the DEPMPO-OOH adduct (x; $a^{\rm N}$ = 13.4 G, $a_{\rm \beta}^{\rm H}$ = 11.9 G, $a^{\rm P}$ = 52.5 G, g = 2.0059) (14), and the carboncentered spin adduct of DEPMPO (‡; $a^{\rm N}$ = 14.5 G, $a_{\rm \beta}^{\rm H}$ = 22.0 G, $a^{\rm P}$ = 46.0 G, g = 2.0061). The assignment of each

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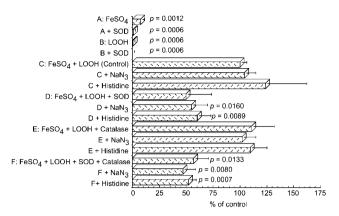


Fig. 3. The effects of SOD (0.5 μ M), catalase (20 μ g/ml), sodium azide (1 mM), and/or histidine (10 mM) on the chemiluminescence observed during the 2 μ M Fe²⁺-catalyzed decomposition of 50 μ M LOOH in the presence of 10 μ M MCLA at 25°C under atomospheric air. The P values were obtained by comparison with a control, using a paired t test.

was confirmed by means of quenching reactions with SOD, catalase and ethanol, as described below. These signals were not observed when DEPMPO was incubated with only Fe²⁺ (Fig. 2B). The addition of SOD reduced the DEPMPO-OOH signal intensity (x; Fig. 2C). The addition of catalase substantially decreased the DEPMPO-OH

signal intensity (o; Fig. 2D). The addition of ethanol to the reaction mixture as an OH scavenger gave the ESR spectrum shown in Fig. 2E. The signals enhanced by the addition of ethanol were identified as those of the DEPMPO adduct of the hydroxyethyl radical [Δ ; DEPMPO-CH(CH₃)OH] ($a^{\rm N}$ = 14.6 G, $a_{\rm p}^{\rm H}$ = 21.1 G, $a^{\rm P}$ = 49.6 G, g = 2.0058; our unpublished data). The ESR parameters of DEPMPO-CH(CH₃)OH are nearly identical to those of the carbon-centered spin adduct of DEPMPO obtained in the decomposition reaction. The present spin trapping experiments suggest the generation of O_2 . OH, H_2O_2 , and the carbon-centered radical during the Fe²⁺-catalyzed decomposition of LOOH under aerobic conditions.

Chemiluminescence during Fe^{2+} -Catalyzed Decomposition of LOOH under Aerobic Conditions—The formation of O_2 — during the present reaction was also examined using MCLA, which is a specific chemiluminescence probe for O_2 — and 1O_2 (24). Fig. 3 depicts the relative intensity of MCLA-dependent chemiluminescence observed during the Fe²⁺-catalyzed decomposition of LOOH under various conditions. The addition of SOD to the control reaction mixture significantly decreased the MCLA-dependent chemiluminescence intensity, suggesting the involvement of O_2 — formation. Since catalase, sodium azide and histidine did not affect the chemiluminescence intensity, hydrogen peroxide (H_2O_2) and 1O_2

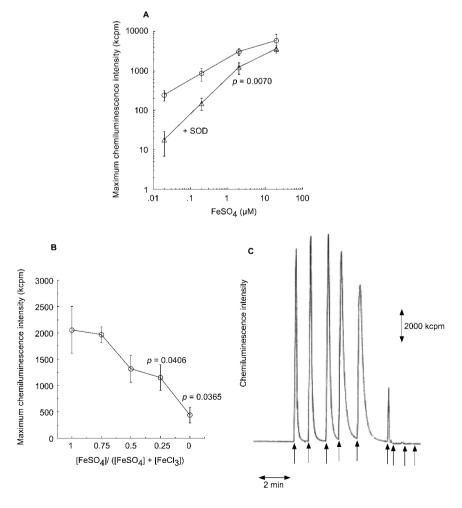


Fig. 4. (A) The effects of the Fe²⁺ concentration on the MCLA-dependent chemiluminescence observed during the ferrous ion-catalyzed decomposition of 50 μM LOOH at 25°C in the presence (Δ) and absence (o) of SOD. The P values were obtained through a paired t test, that compared the results in the presence and absence of SOD. (B) The effect of the ratio between FeSO₄ and FeCl₂ (Fe = 2 µM) on the chemiluminescence intensity served during the iron-catalyzed decomposition of 50 μ M LOOH. The P values were obtained through a paired t test involving comparison with the FeSO₄ (without FeCl₃) system. (C) Chemiluminescence observed in the presence of 10 μM MCLA, 50 µM LOOH, and FeSO₄ in 50 mM potassium phosphate buffer (pH 7.4) at 25°C. 20 µM FeSO₄ was added at each point indicated by an arrow.

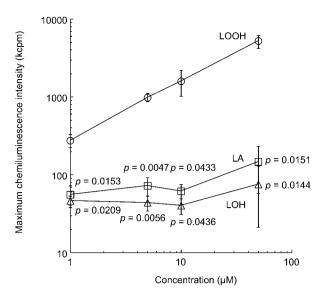


Fig. 5. The effects of the LA, LOH and LOOH concentrations on the MCLA-dependent chemiluminescence observed during the reaction between 2 μM ferrous ion and LA, LOH, and LOOH at 25°C. The *P* values were obtained through a paired t test involving comparison with the LOOH system.

were not contributors to the emission. Judging from the time-dependent change in the LOOH concentration [as monitored by HPLC-CL (18)], the Fe²+-catalyzed decomposition of LOOH in potassium phosphate buffer stopped within an extremely short time (data not shown), probably because Fe²+ was chelated by phosphate and/or converted to ferric hydroxide [Fe(OH)₃], which is poorly soluble in water (1). The addition of DMPO (12 mM) had no effect on the chemiluminescence intensity (data not shown). This was due to the relatively small second-order rate constant of the reaction between O_2 - and DMPO (10 M⁻¹ s⁻¹ at pH 7.8, 25° C) (25) in comparison with that of the reaction between MCLA and O_2 - (2.2×10^5 M⁻¹ s⁻¹ at pH 7, 20° C; K. Akutsu and K. Fujimori, unpublished data).

Strong emission was not observed for the system with MCLA + Fe²⁺ or MCLA + LOOH. The chemiluminescence intensity observed during the decomposition of LOOH by Fe²⁺ was dependent on the FeSO₄ concentration (Fig. 4A). SOD decreased the chemiluminescence intensity but DMPO did not (data not shown). The effect of the ratio between FeSO₄ and FeCl₃ on the chemiluminescence observed during the 2 µM iron-catalyzed decomposition of LOOH was examined (Fig. 4B). When more FeSO₄ was in the reaction mixture, stronger chemiluminescence was observed. Moreover, the addition of 0.1-0.4 µM FeCl₃ to the reaction mixture containing 2 µM FeSO₄, 50 µM LOOH, and 10 µM MCLA had no effect on the chemiluminescence intensity (data not shown). On the other hand, chemiluminescence was observed whenever 20 uM FeSO was added to the solution containing 10 µM MCLA and 50 µM LOOH until LOOH was completely decomposed (Fig. 4C). MCLA was not completely consumed during this experiment (data not shown). These results suggest that FeSO₄, not FeCl₃, was necessary for O₂⁻⁻ production. The intensity of the chemiluminescence was also dependent on the LOOH concentration (Fig. 5). Concentration-

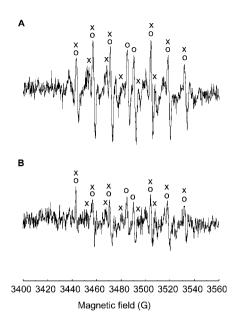


Fig. 6. The effect of argon gas on the DEPMPO-OOH signal intensity obtained during the 100 μM ferrous ion-catalyzed decomposition of 2.2 mM LOOH in the presence of 20 mM DEPMPO at 25°C. ESR spectra were measured at 25°C under atmospheric air. o and x represent the DEPMPO-OH spin adduct and DEPMPO-OOH spin adduct, respectively. A: No treatment, B: Argon gas bubbling. Receiver gain: 5.0 × 10⁵; modulation frequency: 100 kHz; modulation amplitude: 2.018 G; conversion: 81.920 ms; time constant: 40.960 ms; sweep time: 83.886 s; microwave frequency: 9.779 GHz; microwave power: 9.7 mW; scan number: ×20.

dependency of the chemiluminescence intensity was, however, not observed when LA or LOH was used instead of LOOH. Also, an ESR signal was not observed for the system with LA or LOH (data not shown). These results show that LOOH is necessary for the production of O_2 in this system, and hydroperoxide, not carboxylic groupcontaining compounds for chelating iron, is important for the production of O_2 during the Fe²+-catalyzed decomposition of LOOH. Comparable but weak ESR signals and MCLA-dependent chemiluminescence were observed when Fe³+ was used instead of Fe²+ (data not shown).

Yield of O_2 during Fe^{2+} -Catalyzed Decomposition of LOOH—We estimated the approximate ratio of O_2 formed to LOOH decomposed. One count of MCLA-dependent chemiluminescence intensity corresponded to 1 fmol/ml of O_2 formed. The integrated MCLA-dependent chemiluminescence intensity obtained during the 2 μ M Fe^{2+} -catalyzed decomposition of 36 μ M LOOH was 127 \pm 26 kcounts (mean \pm SD, n=3). In the presence of 0.5 μ M SOD, the integrated chemiluminescence intensity was 23 \pm 5 kcounts (mean \pm SD, n=3). The difference was 104 kcounts. This corresponds to 104 pmol/ml of O_2 formed. Since 10% of LOOH was decomposed during the reaction (data not shown), the O_2 formed in this system corresponded to 2.9% of LOOH decomposed.

Effect of Argon Gas in the Solvent on the Intensity of the Spin Adduct Formed during Fe^{2+} -Catalyzed Decomposition of LOOH—Figure 6 shows the effect of argon gas, introduced into the solvent, on the formation of O_2 during the Fe^{2+} -catalyzed decomposition of LOOH. Argon gas bubbled into each solution for 2 h under a nitrogen

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Fe²⁺ + LOOH
$$R_1 - C - R_2$$
 (LO') + 'OH + Fe³⁺

H

OH

R₁ - C - R₂

OO'

R₁ - C - R₂

OO'

 $R_1 - C - R_2$
 $R_1 - C - R_2$
 $R_2 - C - R_2$
 $R_1 - C - R_2$
 $R_2 - C - R_2$
 $R_1 - C - R_2$
 $R_2 - C - R_2$
 $R_2 - C - R_2$
 $R_3 - C - R_2$
 $R_4 - C - R_2$
 $R_4 - C - R_2$
 $R_4 - C - R_2$
 $R_5 - C - R_2$
 $R_7 - C - R_2$

Scheme 1. Proposed mechanism for the formation of the superoxide anion and other active oxygen species during the ferrous ion-catalyzed decomposition of linoleic acid hydroperoxide under aerobic conditions.

atmosphere decreased the ESR signal intensities of DEPMPO adducts (DEPMPO-OOH: $a_{\beta}^{\rm H}=10.7$ G, $a^{\rm N}=13.8$ G, $a^{\rm P}=50.0$ G; DEPMPO-OH: $a_{\beta}^{\rm H}=13.8$ G, $a^{\rm N}=13.6$ G, $a^{\rm P}=46.9$ G). A weak ESR signal was observed in this case since oxygen might remain in the solution and/or the Fe³+-catalyzed nucleophilic attack of water on DEPMPO might occur. The intensity of MCLA-dependent chemiluminescence observed during the Fe²+-catalyzed decomposition of LOOH was also decreased to two-thirds by argon gas bubbling for 30 min under atmospheric air (data not shown).

DISCUSSION

The present study has shown that O_2 -, H_2O_2 , OH, and the carbon-centered radical are formed during the Fe²⁺catalyzed decomposition of LOOH under aerobic conditions. Therefore, DMPO-OH is suggested to be formed through the reaction between DMPO and OH produced via the Fenton reaction. Catalase, however, did not suppress the formation of DMPO-OH and DEPMPO-OH completely. Parts of the observed DMPO-OH and DEPMPO-OH might be produced via other pathways. Koga et al. reported (26) that ¹O₂ is formed during the Fe³⁺-catalyzed decomposition of LOOH, probably via the Russell mechanism (27). The formation of DMPO-OH during the reaction between DMPO and ¹O₂ has also been reported (10, 11). However, no effect of an ¹O₂ quencher (sodium azide, uric acid, or histidine) on the ESR signal (data not shown) or chemiluminescence intensity was observed in the present study. Schaich and Borg reported that the spectrum of the DMPO-lipid alkoxyl radical was identical to that of DMPO-OH (4), but this appears not to be correct since the DMPO-linoleic acid alkoxyl radical spin adduct spectrum reported recently (28) was different from the DMPO-OH spectrum.

The main radical species formed during the Fe²⁺-catalyzed decomposition of LOOH is LO \cdot (1, 2). Therefore, MCLA may react with LO \cdot , the MCLA radical may be formed, and light may be emitted, although MCLA is known as a specific probe for the chemiluminescent detection of O₂ $^-$ and 1 O₂ (24). Since the reaction between the MCLA radical and molecular oxygen is significantly

slower than that of dimerization of the MCLA radical (29), MCLA does not react with oxygen to emit light in the present system. Therefore, the MCLA-dependent chemiluminescence observed in this study was caused by O_2 .

Since each ESR spectrum was obtained by the accumulation of ten spectra obtained consecutively (each scan took 83.883 s), measurement was continued after the reaction had been stopped. On the other hand, the half-life of DMPO-OOH is 60 s (25) and that of DEPMPO-OOH is 890 s (14). Therefore, DMPO-OOH decomposed into DMPO-OH during the measurement and DEPMPO-OOH remained. For these reasons, the DEPMPO-OOH spin adduct, but not the DMPO-OOH one, might be observed during the Fe²⁺-catalyzed decomposition of LOOH.

The present study has shown that LOOH was needed for the formation of O_2^{-} during the decomposition of LOOH by Fe^{2+} . Our results also show that Fe^{2+} was needed for O_2^{-} formation. The Fe^{2+} -catalyzed reduction of triplet oxygen to O_2^{-} (1) contributed only slightly to the formation of O_2^{-} in the present study (Figs. 1–3). On the other hand, introducing argon gas into the solvent reduced the intensity of both the ESR signal (Fig. 6) and the MCLA-dependent chemiluminescence (data not shown) observed during the Fe^{2+} -catalyzed decomposition of LOOH. These results suggest that oxygen dissolved in the solution, in addition to the reaction intermediate generated during the decomposition of LOOH, plays an important role in the production of O_2^{-} in the present reaction system.

Recently, it was suggested that primary alkoxyl radicals (RCO+) are converted into α-hydroxylcarbon-centered radicals through an unusual 1,2-H-atom shift in the presence of water (30), and the carbon-centered radicals react with dioxygen to yield α-hydroxyperoxyl radicals (30, 31). It is well known that a number of peroxyl radicals decompose into carbocations and O_2 . Therefore, we tentatively propose the mechanism of O_2 formation shown in Scheme 1. Conversion of LO [R₁CH(O+)R₂] into a hydroxycarbon-centered radical [R₁C⁺ (OH)R₂] in water may occur, although it is a secondary alkoxyl radical. In fact, a carbon-centered radical was detected during the Fe²⁺-catalyzed decomposition of LOOH under anaerobic conditions (32). In the present spin trapping experiments involving DEPMPO, the carbon-centered radical was also detected during the reaction. Then, the hydroxyperoxy radical [R₁COO (OH)R₂] formed through the reaction between this carbon-centered radical and molecular oxygen may decompose into a carbocation [R₁C⁺(OH)R₂] and O_2 .

Finally, in this study we have demonstrated for the first time the mechanism of formation of O_2 . (as distinct from the Fe²⁺-catalyzed reduction of oxygen) during the Fe²⁺-induced decomposition of LOOH under aerobic conditions.

We wish to thank Dr. Yasuhiro Kobori of Tohoku University for the advice to Y.K. regarding the ESR technique.

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